

Anatomical study with antibacterial and antibiofilm effects of *Erodium cicutarium* (L.) L'H phenolic roots extract

Zubaida A. Lattaf Ismaeel

Department of Biology, AL-Iraqia University, Baghdad, Iraq

E-mail: dr.zubaidalattaf@gmail.com

ABSTRACT

Bacteria acquire resistance to antibacterial drugs, so there is always a need for new drugs to inhibit many human bacterial pathogens. Novel studies were carried out to examine the internal structures as well as the antibacterial effect of *Erodium cicutarium* phenolic root extracts (APR) on the development of biofilm formation of bacterial species, *Klebsiella pneumoniae* and *Staphylococcus aureus* using four strains from both clinical and environmental samples *in vitro*. Anatomy of all the vegetative parts showed a presence of tannin-filled cells in their structures. Also, the experiments were conducted to study the antibacterial effect by determining the minimum inhibitory concentrations (MICs), and susceptibility test of these strains for five antibiotics. Although the APR concentrations of 50, 25, 12.5, 6.25, 3.125, and 1.625 $\mu\text{g mL}^{-1}$; W/V) were used through antibiotic and biofilm inhibition assay. The result of sensitivity test showed more resistant to various antimicrobial especially Amoxicillin, Cephalothin and Methicillin for both bacterial strains. In addition, the MICs tested showed that the ranged values were in concentrations between 0.017 to 0.650 mg mL^{-1} for both *K. pneumoniae* and *S. aureus* strains. Furthermore, biofilm reduction assay results for treatments showed that the highest activity was obtained with phenolic root extracted, biofilm eradication at 1.3 mg mL^{-1} . In conclusion, the results showed that relatively low concentrations of phenolic root extracts displayed promising antibacterial and antibiofilm capabilities making them attractive for additional studies as “novel therapeutic agents.”

Keywords: Anatomical, Antibacterial, Antibiofilm, *Erodium*, *Klebsiella*, *Staphylococcus*.

Article type: Research Article.

INTRODUCTION

Geraniaceae is Cosmopolitan family of mostly temperate and subtropical annual and perennial herbs and a few small shrubs, comprising about 750 species belong to 11 genera. There are 74 species belonging to the genus *Erodium*, which can be found in many regions, especially in the Mediterranean basin (Fiz *et al.* 2006). The most common of these species is *E. cicutarium*. It was annual and naturally self-growing five-leaved herbaceous plant with pink blooms in spring (Latimer *et al.* 2019; Pieroni & Cattero 2019). Although this species is consumed as fresh and dried vegetable in the regions where it grows, it is widely used for traditional medicine (Duke 2001). It is used as a traditional medicine in many diseases, especially in diseases such as dysentery, uterine bleeding, constipation and diabetes (De la Cruz *et al.* 2007; Tene *et al.* 2007; Molares & Ladio 2009; Özgün *et al.* 2012; Safa *et al.* 2012; Rajaei & Mohamadi 2012; Asadi Samani *et al.* 2016; Munekata *et al.* 2019). The fresh form of the plant is mostly used as a vegetable, while the dry form of the plant as a traditional medicine. The principal active ingredients of *Erodium* include Tannin, catechins, gallic and elagic acids, sugars (glucose, galactose and fructose), amino acids (glycine, alanine, proline, histidine, tryptophan, tyrosine and glutamic acid), vitamins K and C (Al Snafi 2017). Plant anatomy, the study of the internal structure of plants, had been a source of fascination and field of scientific inquiry since the time of earliest microscopists. It plays an important role in the understanding of plant biology. A realistic interpretation of morphology, physiology and phylogeny should be based on a thorough knowledge of cells structure and tissue (Dengler 2002; Hamad *et al.* 2023; Ahmed *et al.*

2023; Iriani *et al.* 2023). Among the factors contributing to microbial resistance is the ability of the microbes to exist in biofilm forms that allow them to withstand harsh environmental conditions and antimicrobial agents. *Staphylococcus aureus* is the most common infectious agent involved in the development of skin infections that are associated with antibiotic resistance, such as burn wounds. Also, *Klebsiella pneumoniae* is the second most common cause of gram-negative bacteriemia after *Escherichia coli*. Bacteriemia causes significant morbidity and mortality in general populations (Lee 2003; Tsay *et al.* 2002). Biofilms can be very problematic in various aspects of our lives ranging from medical to industrial areas. In addition to their increased resistance to antimicrobial agents, biofilms can form on many medical implants such as catheters, artificial hips, and contact lenses. The most worrisome fact is that cells existing in a biofilm can become 10-1000 times more resistant to antimicrobial agents, mainly through the production of extracellular polymeric substance matrix that hinders the access of antibiotics to the bacterial cells. These infections can often only be treated by removal of the implant, thus increasing the trauma to the patient and the cost of the treatment. It has been estimated that biofilms are associated with 65% of nosocomial infections and that treatment of these biofilm-based infections costs > \$1 billion annually (Cox & Balick 1994). The aim of this study was to identify anatomical characters of root, stem, petiole and leaf blades of *E. cicutarium* in addition to evaluate the antibacterial effect of phenolic root extracts (APR) and on the development of biofilm formation of bacterial species such as *K. pneumoniae* and *S. aureus* by using four strains.

MATERIALS AND METHODS

Collection and extract preparation of the studied plant

Plant materials were collected from various parts of Mosul, Iraq during the flowering period in 2022. Authentication of the plant was carried out at the herbarium of the Department of Biology, College of Science, Baghdad University, Iraq. Afterward, the plant materials were rinsed thoroughly with tap water to remove extraneous contaminants. The wanted parts for the anatomical study were slit into small pieces and fixed in formalin acetic acid solution. After 24 h, the plant parts were washed several times with ethyl alcohol solution with concentration of 70% then saved in it even use (Ismaeel 2022). The method of AL Musawi (1979) and AL Zubaidy (1998) were used with some modifications. Some sections stained by safranin and glycerine mixture (1:10). For extract preparation, roots were cut into small pieces, oven-dried at 50 °C until stability of dry weight was observed, and then grounded into powder with an electric-grinder to prepare it for extraction (Harborne 1984).

Crude phenols extract

Phenolic compounds separation

The separation of the phenolic component was performed according to (Harborne 1984). Approximately 200 g of root powder was shaken with 1 L of 80% ethanol for 72 h in cool and dark location. Then the extract was filtered and dried at a temperature of 30-40 °C via a rotary evaporator. For acid hydrolysis 10% concentration HCl was used for 10- 30 min in a water bath. This step resulted in the hydrolyse the glycosidic linkage to get aglyconic part, cooled and filtrated. Finally, the mixture was extracted using chloroform (1:1) in three times separation fennel. The polyphenolic fraction (chloroform layer) was collected and submitted to rotary evaporator to remove the solvent form. The ferric chloride test was applied to detect phenol compounds in the row extract. Then different concentrations of the extract were prepared (0.3, 0.5 and 1.3 mg mL⁻¹).

Microorganisms

The bacteria *K. pneumoniae* and *S. aureus* strains were obtained from Microbiology Laboratory in the College of Sciences, University of Baghdad. The strains were sub-cultured into cetrinide agar plates and incubated at 37 °C for 24 h. Following incubation, single colonies were transferred from the plates and inoculated into tubes containing brain heart broth. The cultures were incubated at 37 °C for 24 h before used (Kaya *et al.* 2020).

Antibiotic susceptibility test

From an overnight culture plate, 4-5 colonies of *K. pneumoniae* and *S. aureus* strains were picked up and suspended in 5-mL sterile normal saline until the turbidity was a proximately equivalent to that of the McFarland No. 0.5 turbidity standard. Ten minutes later, by a sterile forceps the antimicrobial disc including Amoxicillin, Ampicillin, Cefotaxime, Cephalothin and Methicillin were picked up and placed on the surface of Mueller Hinton plates (OECD 2001). The plates were incubated at 37 °C for 18-24 h. After incubation, the plates were examined

for the presence of inhibition zone of bacterial growth around the antimicrobial discs. The diameter of the zone of inhibition was measured by mL using a metric ruler and compared to standard inhibition zone according to Clinical and Laboratory Standards Institute (OECD 2008).

Determination of minimum inhibitory concentration (MIC)

MIC of plant extracts was determined by microdilution method in sterile 96-wells microliter plates according to the protocol described previously (McBain *et al.* 2004). Different plant extracts concentrations (50, 25, 12.5, 6.25, 3.125 and 1.625 $\mu\text{g mL}^{-1}$; W/V) were prepared containing bacterial cells comparable to McFarland standard no. 0.5 in a final volume of 200 μL . Sterile distilled water, broth and plant extracts were used as a negative control while sterile distilled water, broth and bacteria positive control. After 24 h at 37 °C, the MIC of each sample was determined. The MIC was considered as the lowest concentration of an antimicrobial that will inhibit the visible growth of a microorganism after 24 h incubation.

Effect on adherence and biofilm formation

The effect of different concentrations of plant extracts and on adherence and biofilm-forming ability was tested on polystyrene flat-bottomed microtiter plates as described by (Saising *et al.* 2012) with some modifications. Overnight cultures were diluted (1:100) with tryptone soy broth supplemented with 1% (w/v) glucose. (200 μL) from the culture were then transferred to the wells of a 96-well polystyrene microliter plate and incubated overnight at 37 °C. After incubation, supernatant part was removed from each well and the plate were gently washed twice with normal saline, then dried and fixed at 65 °C for 1 h. Therefore, all the plates were stained with 0.1% (w/v) crystal violet for 10 min, gently washed and the quantitative analysis of biofilm was performed by adding 200 μL 95% ethanol for 10 min. Finally, the biofilm was measured at 630 nm by microplate reader in the present of the methylene blue in the de-staining solution (ethanol).

Statistical Analysis

Complete Randomized Design (CRD) was used as experimental design. Data were analysed using statistical analysis system- SAS to study the effect of different plant extracts and the nanoparticles on some bacterial isolates. Least significant difference (LSD) was used to compare the significant difference between means. Differences were considered significant when $P \leq 0.05$ (Mason *et al.* 2003).

RESULTS AND DISCUSSION

Anatomical study

In transverse section of *E. cicutarium* roots, the periderm layer on the outermost surface was rectangular to cuboidal cells arranged in 4-6 rows with mean thickness of 58.02 μm . The multilayered parenchymatous cortex composed of numerous of storage parenchyma cells and characterized by presence of tannin filled cells in the tissue. Mean thickness of the cortex layer was 121.66 μm . The phloem seemed as a conical masses of cells attached from the base as a continuous ring with mean thickness 21.36 μm . Xylem was regular diffused porous wood with mean half diameter of 413.33 μm (Plate 1 A & B). In transverse sections of the hairy stem of the studied species, shape of the stem was circular. Epidermis was single layered covered by thin layer of minute projections cuticle with mean thickness of 5.93 μm . Epidermal cells were ovate to oblong ovate, thickened vary from 13.6 μm to 21.76 μm . Below the epidermis, cortex represented by 13-16 rows of ovate to oblong ovate parenchymatous cells with mean thickness of 168.41 μm containing starch grains and tannin filled cells in highly amounts. Pericycle contained a well defined continuous one ring of sclerenchyma, which comprised of unite of the arcs of interfascicular sclerenchyma with the fibre caps of the bundles, so as to form a continuous one ring. The caps of the pericyclic fibres were smaller in diameter than those of the inter-fascicular sclerenchyma arcs. Central cylinder composed of 13-15 collateral vascular bundles separated by medullary rays. Wood had crescent shape and the tracheary elements arranged as groups. Pith consisted of circular to sub circular parenchymatous cells. These cells stored highly amounts of starch in addition to tannic acid filled cells as clusters in scattered regions (Plate 1 C&D). Transverse sections of the hairy leaf showed that, it was covered with one layer of epidermis with polygonal or rectangular ordinary epidermal cells with thickness of 24.48 μm to 27.74 μm . The outer walls of the epidermal cells hade well developed cuticle of 3.98 μm thick. The palisade region of the leaf consisted of 3 – 4 rows of elongate – prismatic shaped cells, then spongy layer beneath the palisade layer comprised of 3 – 4 rows of nearly

isodiametric cells. One central circular –ovate vascular bundle was observed in leaf centre. Tracheary elements of the bundle were arranged as group. In the midrib of the leaf, large amount of tannin filled cells were observed (Plate 1 E). Cross sections of petioles had crescent shape comprised of the outermost layer; cuticle, with mean thickness of 6.8 μm covering the uniseriate epidermis which composed of oblong - ovate to sub-rectangular cells with mean thickness of 19.22 μm . Cortex was parenchymatous tissue type with the outer rows specialized as chlorenchyma. Petiole had five conical vascular bundles with tracheary elements arranged in form of group. Phloem groups were bounded externally by caps of fibres. Petioles tissue had tannin filled cells, storage parenchyma cells and druses crystals (Plate 1 F&G). Tannin-filled cells were observed in all the studied parts. This result showed a parallelism with the record of (Metcalf & Chalk 1950; Al Mayah & Hammadi 1998; Al Helfy 2000).

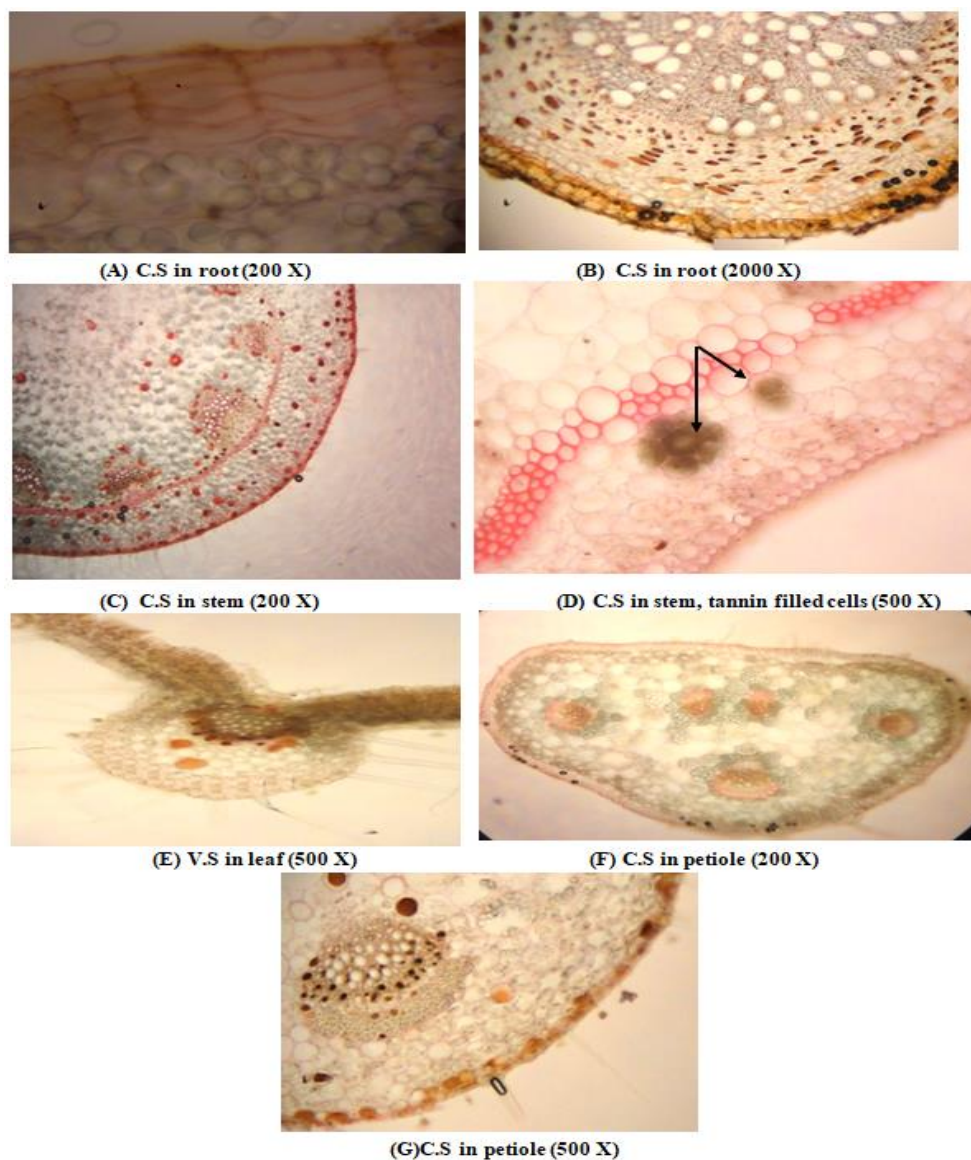


Plate 1. Anatomical sectioning of root, stem, leaf and petiole of *E. cicutarium*

Bacterial susceptibility to antibiotics

In this study, the bacterial strains were chosen because of the importance of these strains in the hospital environment and their outbreak in the community. Susceptibility tests were summarized in Tables 1 and 2 for *S. aureus* and *K. pneumoniae* strains to five different antibiotics by disc diffusion method recommended by CLSI (CLSI 2013). The result of the antibiotic susceptibility tests showed that the bacteria were more resistant to various antimicrobial treatments. These antibiotics were used in this study due to their mode of action inhibiting cell wall synthesis which caused the release of the bacterial cell DNA into the surroundings (Seguin *et al.* 1999).

Table 1 and Table 2 indicate that all strains were resistant to Ampicillin, Amoxicillin. Besides, all *S. aureus* strains were ranging from intermediate and sensitive to Cefotaxime, While the *K. pneumoniae* strains were intermediately resistant to Cefotaxime with more resistance to other antimicrobial drugs.

Table 1. Antibiotic susceptibility of *S. aureus* strains according to CLSI

<i>S. aureus</i> Strains	Amoxicillin	Ampicillin	Cefotaxime	Cephalothin	Methicillin
1	R	R	S	R	R
2	R	R	I	R	R
3	R	R	S	R	R
4	R	R	S	R	R

Note: R = Resistant, I = Intermediate, S = Sensitive.

Table 2. Antibiotic susceptibility of *K. pneumoniae* strains according to CLSI

<i>K. pneumoniae</i> Strains	Amoxicillin	Ampicillin	Cefotaxime	Cephalothin	Methicillin
1	R	S	I	R	R
2	R	S	I	R	R
3	R	R	I	R	R
4	R	R	I	R	S

Note: R = Resistant, I = Intermediate, S = Sensitive.

Multidrug resistance has been recognized as a virulence factor of great scale in clinical infections. Because of the increase in the intricacy of the majority of microbial infections and the resistance to straight treatment, researchers have been prompted to identify alternatives for the action of infections. The plant extracts with biologically active compounds isolated from plants have gained extensive attention in this look as they have been known to cure diseases and sickness since very old times. Silver nanoparticles are also able to assess human health through a variety of commercial products. Studies have shown that silver nanoparticles cause toxicity to germline stem cells through a reduction in mitochondrial function and induction of membrane leakage and apoptosis (AL Shahwany *et al.* 2016).

Evaluation of MIC values

Minimum inhibitory concentration (MIC)

Minimum inhibitory concentration (MIC) value is important to determine efficacy of antibacterial agent. Low MIC value may be an indication of high efficacy or that microorganism has no potential to develop resistance towards the bioactive compound. The result in Table 3 and Figs. 1, 2, 3 and 4 showed that the bacterial strains behaved significant differences in their sensitivity to the different extracts added to their growth medium. Obviously, MIC ranged value was in concentration between 0.017 to 0.650 mg mL⁻¹ for both *K. pneumoniae* and *S. aureus* strains. Moreover, results in Fig. 1 indicated that the *K. pneumoniae* strains were intermediate effects than *S. aureus* strains which were sensitive to APR extracts concentrations. Also, the result in Fig. 2 showed that the lowest MIC was 0.079 with a high APR concentration of 1.3 mg mL and the highest MIC was 0.308 after treatments with a lower APR concentration of 0.3 mg mL for both bacterial strains.

Table 3. Antibacterial activity of *E. cicutarium* root phenolic (APR) extracts against some of *S. aureus* and *K. pneumoniae* strains.

strains	Concentrations						Average	
	0.3 mg mL ⁻¹		0.5 mg mL ⁻¹		1.3 mg mL ⁻¹		Bact. 1	Bact. 2
	Bact. 1	Bact. 2	Bact. 1	Bact. 2	Bact. 1	Bact. 2		
St. 1	0.133	0.273	0.107	0.107	0.120	0.017	0.120	0.132
St. 2	0.133	0.330	0.130	0.170	0.150	0.067	0.138	0.189
St. 3	0.133	0.650	0.107	0.130	0.067	0.080	0.102	0.287
St. 4	0.160	0.650	0.109	0.210	0.068	0.067	0.111	0.309
Average	0.140	0.476	0.113	0.154	0.101	0.058	0.118	0.229
L.S.D 5%				0.018				0.026
					0.037			

Bact. 1: *K. pneumoniae*, Bact. 2: *S. aureus*, St.: strains

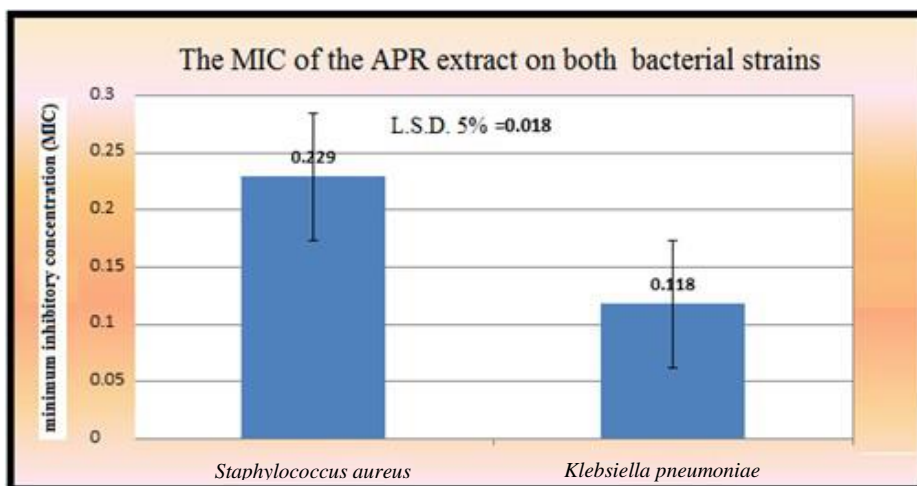


Fig. 1. Minimum inhibitory concentration (MIC) value for *S. aureus* and *K. pneumoniae*.

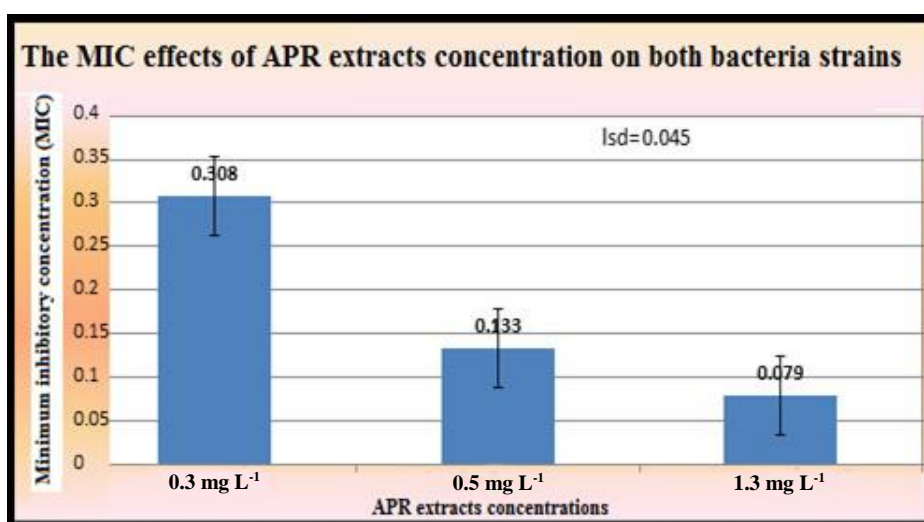


Fig. 2. Minimum inhibitory concentration (MIC) value for APR extracts concentration.

Moreover, the evaluation of the inhibition potential of plant extracts against *S. aureus* and *K. pneumoniae* strains (Figs. 3 and 4) showed that APR extracts present a greater effect on bacterial strains No. 3 and 4 than 1 and 2 for both kinds, especially *S. aureus* strains.

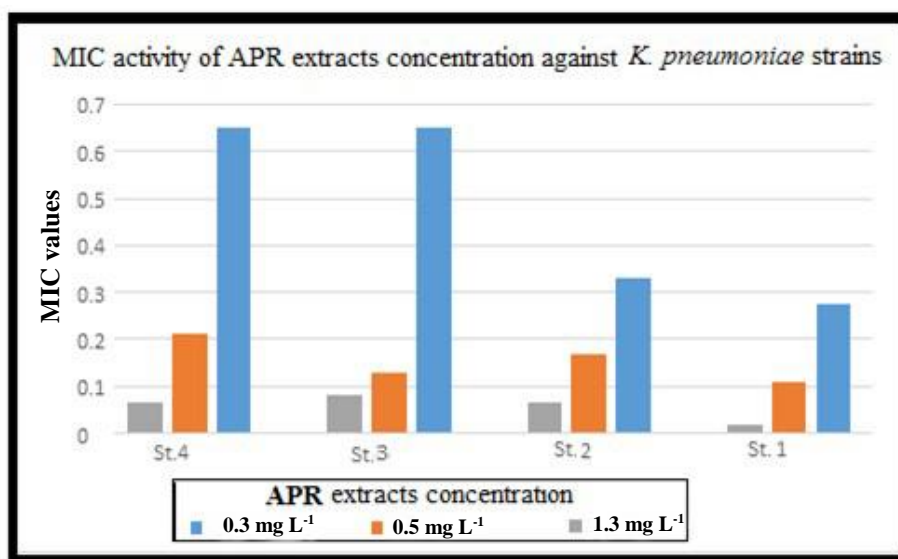


Fig. 3. the interaction effects between Minimum inhibitory concentration (MIC) value for *K. pneumoniae* and the APR extracts concentrations.

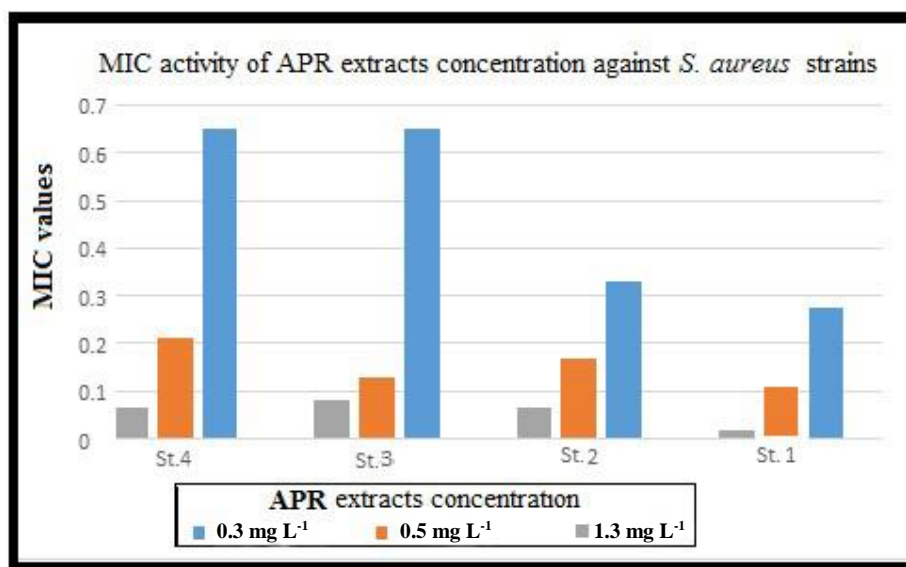


Fig. 4. the interaction effects between Minimum inhibitory concentration (MIC) value for *S. aureus*. And the APR extracts concentrations.

Analysis of the experimental data revealed that organic extracts were more effective against Gram-positive than Gram-negative bacteria. Gram-negative bacteria are highly resistant, and this resistance is likely related to the nature of their outer membranes, which are impervious to lipophilic compounds (Djenane *et al.* 2012). Gram positive bacteria are more sensitive and less protected against polyphenolic agents because they only have an outer layer of peptidoglycans, which can only prevent the diffusion of molecules whose molar mass is greater than 50 000 D (Abirami *et al.* 2012). (Lazar 2011) obtained similar results to those of the present study, and supporting the hypothesis that Gram-positive bacteria are more sensitive to plant extracts. In a recent study, reported that the antimicrobial activity of an extract is likely due to the presence of synergy among various phenolic components.

Antibiofilm activity in a 96-well microplate

Tables 4-5 and Fig. 5 depict significant differences between two bacteria types and plant extract concentrations. The results of *in vitro* biofilms were presented in Tables 4 - 5. Furthermore, biofilm reduction assay results showed that APR at 0.347 mg mL⁻¹ could inhibit 100% of *K. pneumoniae* biofilm and at 0.053 decrease 50 % of biofilm formation (Table 4).

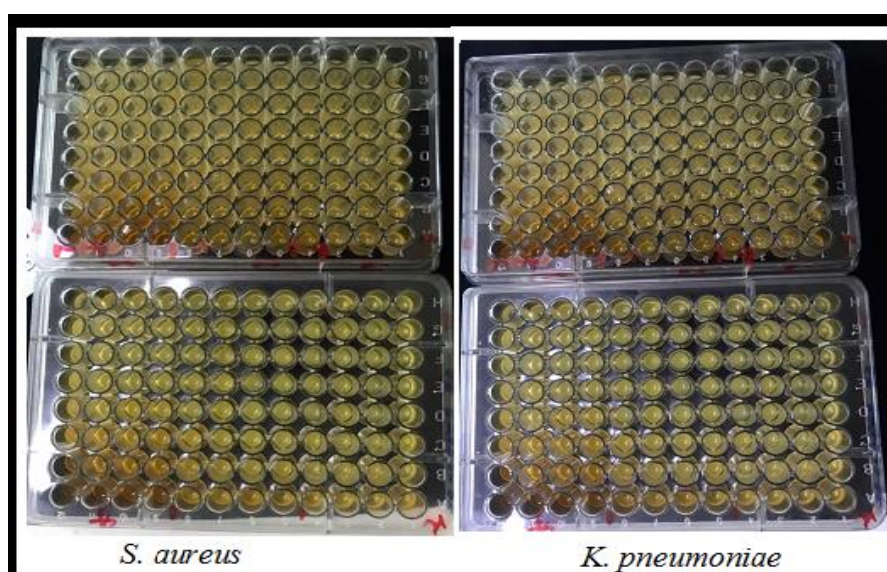


Fig. 5. Biofilm of *S. aureus* and *K. pneumoniae* formation tests.

Furthermore, biofilm reduction assay results showed that the average treatments which decrease significantly biofilm formation for *K. pneumoniae* strains No. 1, 2, 3 and 4 were 0.332, 0.214, 0.154 and 0.192 mg mL⁻¹, respectively, while the average treatments which decrease biofilm formation for *S. aureus* strains No. 1, 2, 3 and 4 were 0.169, 0.285, 0.404, 0.331 mg mL⁻¹, respectively. Noteworthy, significantly the best plant extract effective concentrations was 1.3 mg mL⁻¹ which was able to inhibit biofilm formation by average treatments 0.221 and 0.136 mg mL⁻¹ of all *S. aureus* and *K. pneumoniae* strains, respectively (Tables 4 - 5). The significantly difference in biofilm thickness results from different reasons such as differences in strains capacity to form biofilm, perhaps the primary number of cells that succeeded in adherence and the differences in the quality and quantity of autoinducers (Quorum sensing signaling molecules) that were produced from each strain and played an essential as well as important role in biofilm formation (Lazar 2011).

Table 4. Effects of APR extract on *K. pneumoniae* biofilms formation at 24 h.

Strains	Treatments				Average treatments
	Before	Conc. 0.3 mg mL ⁻¹	Conc. 0.5 mg mL ⁻¹	Conc. 1.3 mg mL ⁻¹	
Iso 1	0.505	0.284	0.344	0.194	0.332
Iso 2	0.367	0.209	0.162	0.119	0.214
Iso 3	0.261	0.086	0.168	0.101	0.154
Iso 4	0.253	0.188	0.198	0.130	0.192
Average treatments	0.347	0.192	0.218	0.136	0.223
L.S.D 5%		Strains 0.008**		Treatments 0.008**	Interaction 0.016**

Table 5. Effects of APR on *S. aureus* biofilms formation at 24 h.

Strains	Treatments				Average treatments
	Before	Conc. 0.3 mg mL ⁻¹	Conc. 0.5 mg mL ⁻¹	Conc. 1.3 mg mL ⁻¹	
Iso 1	0.252	0.233	0.063	0.126	0.169
Iso 2	0.394	0.232	0.291	0.223	0.285
Iso 3	0.680	0.346	0.306	0.283	0.404
Iso 4	0.472	0.225	0.376	0.250	0.331
Average treatments	0.449	0.259	0.259	0.221	0.297
L.S.D 5%		Strains 0.007**		Treatments 0.007**	Interaction 0.013**

CONCLUSION

Increasing drug resistance to traditional treatments observed in several bacterial strains, in addition to harmful APR extracts and substantial cost of treatment shifted the pace toward finding novel therapeutic agents, natural products such as plants and plant-derived compounds were the primary target due to their efficacy, safety, and lower cost. In this study, we selected *E. cicutarium* in an effort to find novel agents that could aid in fighting bacterial infections and their biofilms. MIC values indicated that not all concentrations exhibited an activity. Regarding the MIC assay, only 1.3 mg mL⁻¹ APR extract concentration was able to inhibit bacterial growth, among which was most significant where it achieved the lowest MIC and had an effect against multiple strains. We also noticed that one of our tested strains, *K. pneumoniae* No. 1 was insensitive to all treatments used, i.e., independently of the strain being a Gram-positive one, but rather probably related to the strain itself or the nature of our extract concentration. Furthermore, it was evident that most of the active extracts were thymol, which are characterized by containing flavonoids and phenols that are known antimicrobial agents (Al Snafi 2017). This could be due to a potential synergistic effect between the different phytochemical constituents found in the whole plant.

The synergism effect in plant extracts was also highlighted in literature as in the study of (Ncube *et al.* 2012) on the antimicrobial synergism within plant extract combinations from three South African medicinal bulbs. The plants extract compound (phenolic compound) may provide a safe and highly effective alternative to commonly used antibiotics, which are ineffective towards the antibiotic-resistant *S. aureus* and *K. pneumoniae*. Thus, further studies are required to test their activity against other pathogenic bacteria and fungi and study the possibility of using these active components by drugs companies. Finally, further studies using in vivo models are needed to study the impact of APR on health.

REFERENCES

- Abirami, P, Gomathinayagam, M & Panneerselvam, R 2012, Preliminary Study on the antimicrobial activity of *Enicostemma littorale* using different solvents. *Asian Pacific Journal of Tropical Medicine*, 5: 552-555.
- Ahmed, QA, Abdullah, SI, Taher, HM 2023, Effects of the alcoholic extract of ginseng roots and carob fruits in comparison with vitamin E in improving the efficiency of the male reproductive system of albino rabbits. *Caspian Journal of Environmental Sciences*, 21: 815-826.
- Al Helfy, MAA 2000, Systematic study of the genus *Erodium* in Iraq. PhD Dissertation, University of Basrah, Iraq.
- Al Mayah, AA & Hammadi, KJ 1998, Vegetative anatomy of *Polygonum* (Polygonaceae). *Basrah Journal of Science*, 16: 55-62.
- AL Musawi, AH 1979, A systematic study of the genus *Hyoscyamus* (Solanaceae). PhD Dissertation, University of Reading, UK, 96 p.
- AL Shahwany, AW, Tawfeeq, HK & Hamed, ShE 2016, Antibacterial and anti-biofilm activity of three phenolic plant extracts and silver nanoparticles on *Staphylococcus aureus* and *Klebsiella pneumoniae*. *Biomedicine and Biotechnology*, 4: 12-18.
- Al Snafi, AE 2017, A review on *Erodium cicutarium*: A potential medicinal plant. *Indo American Journal of Pharmaceutical Research*, 4: 110-116.
- Al Snafi, AE 2017, Therapeutic potential of *Erodium cicutarium*: A Review. *Indo American Journal of Pharmaceutical Research*, 4:407-413.
- AL Zubaidy, AMA 1998, Systematic study of the genera (*Ajuga* L., *Marrubium* L., *Lallemanita* Fisch. and C.A. Mey and *Laminm* L.) of Labiatae in Iraq. PhD Dissertation, University of Baghdad, Iraq.
- Asadi Samani, M, Moradi, MT, Bahmani, M & Shahrani, M 2016, Antiviral medicinal plants of Iran: a review of ethnobotanical evidence. *International Journal of PharmTech Research*, 9: 427-434.
- CLSI 2013, Clinical and Laboratory Standards Institute. <http://clsi.org/membership/current/my-clsi/>.
- Cox, PA & Balick MJ 1994, The ethnobotanical approach to drug discovery. *Scientific American*, 270: 82-87.
- De la Cruz, H, Vilcapoma, G & Zevallos, PA 2007, Ethnobotanical study of medicinal plants used by the Andean people of Canta, Lima, Peru. *Journal of Ethnopharmacology*, 111: 284-294.
- Dengler, NG 2002, An integral part of Botany (Book review). *American Journal of Botany*, 89: 369-374.
- Djenane, D, Yangüela, J, Derriche, F, Bouarab, L & Roncales P 2012, Use of olive leaf compounds as antimicrobial agents: Application for preserving fresh turkey meat. *Natural Technology*, 7: 53-61.
- Duke, JA 2001, (Ed.) Handbook of Edible Weeds. CRC Press, Boca Ratón, FL.
- Fiz, O, Vargas, P, Alarcon, ML & Aldasoro, JJ 2006, Phylogenetic relationships and evolution in *Erodium* (*Geraniaceae*) based on trnL-trnF sequences. *Systematic Botany*, 31: 739-763.
- Hamad, MH, Hadi, ME & Alsaffar, MF 2023, Effects of some alcoholic extracts of propolis in ovulation and fertility rate of the ovary and oviduct in quail. *Caspian Journal of Environmental Sciences*, 21: 317-323.
- Harborne, JB 1984, *Phytochemical methods*. Chapman and Hall. New York, 2nd ed., 288 pp.
- Iriani, F, Rakhmiati, R, Kertasari, VD, Handayani, EP, Zuyasna, Z, W. Yanuastri, P 2023, Secondary metabolite compounds of *Moringa oleifera* leaves in two different urban altitude locations, Indonesia. *Caspian Journal of Environmental Sciences*, 21: 1065- 1071.
- Ismaeel, ZAL 2022, Comparative anatomical study for some wild species belong to Amaranthaceae and Compositae in Iraq. *Biomedical and Pharmacology Journal*, 15: 1737-1743.
- Kaya, C, Higgs, D, Ashraf, M, Alyemini MN & Ahmad, P 2020, Integrative roles of nitric oxide and hydrogen sulfide in melatonin-induced tolerance of pepper (*Capsicum annuum* L.) plants to iron deficiency and salt stress alone or in combination. *Physiologia Plantarum*, 168: 256-277.
- Latimer, AM, Jacobs, BS, Gianoli, E, Heger, T & Salgado Luarte, C 2019, Parallel functional differentiation of an invasive annual plant on two continents. *AoB Plants*, 11: 1-16.
- Lazar, V 2011, Quorum sensing in biofilms: How to destroy the bacterial citadels or their cohesion/power. *Anaerobe*, 17: 280-285.
- Lee, JH 2003, Methicillin (Oxacillin)-Resistant *Staphylococcus aureus* strains isolated from major food animals and their potential transmission to humans. *Applied and Environmental Microbiology*, 69: 6489-6494.
- Mason, RL, Gunst, RF & Hess, JL 2003, Statistical design and analysis of experiments. Welly-Interscience, New Jersey.

- McBain, AJ, Ledder, RG, Srinivasan, P & Gilbert, P 2004, Selection for high-level resistance by chronic triclosan exposure is not universal. *Journal of Antimicrobial Chemotherapy*, 53: 772-777.
- Metcalf, CR & Chalk, L 1950, Anatomy of Dicotyledons. Vol. 1. Clarendon press. Oxford. PP. 1500.
- Molares, S & Ladio, A 2009, Ethnobotanical review of the Mapuche medicinal flora: Use patterns on a regional scale. *Journal of Ethnopharmacology*, 122: 251-260.
- Munekata, PES, Alcántara, C, Collado, MC, Garcia Perez, JV, Saraiva, JA, Lopes, RP, Barba, FJ, do Prado Silva, L, Sant'Ana, AS, Fierro, EM & Lorenzo, JM 2019, Ethnopharmacology, phytochemistry and biological activity of *Erodium* species: a review. *Food Research International*, 126: 108659.
- Ncube, B, Finnie, JF & Van Staden J 2012, *In vitro* antimicrobial synergism within plant extract combinations from three South African medicinal bulbs. *Journal of Ethnopharmacology*, 139: 81-9.
- OECD 2001, Guidelines for testing of chemicals. Acute oral toxicities up and down procedure, OECD Publishing, 425: 1-26.
- OECD 2008, Test no. 425: Acute oral toxicity: Up-and-down procedure, OECD Publishing.
- Özgen, U, Kaya, Y & Houghton, P 2012, Folk medicines in the villages of Ilica District (Erzurum, Turkey). *Turkish Journal of Biology*, 36: 93-106.
- Pieroni, A & Cattero, V 2019, Wild vegetables do not lie: comparative gastronomic ethnobotany and ethnolinguistics on the Greek traces of the Mediterranean diet of southeastern Italy. *Acta Botanica Brasilica*, 33: 198-211.
- Rajaei, P & Mohamadi, N 2012, Ethnobotanical study of medicinal plants of Hezar mountain allocated in south east of Iran. *Iranian Journal of Pharmaceutical Research*, 11: 1153-1167.
- Safa, O, Soltanipoor, MA, Rastegar, S, Kazemi, M, Dehkordi, KN & Ghannadi, AR 2012, An ethnobotanical survey on Hormozgan Province, Iran. *Avicenna Journal of Phytomedicine*, 3: 64-81.
- Saising, J, Dube, L, Ziebandt, AK, Voravuthikunchai, SP, Nega, M & Götz F 2012, Activity of Gallidermin on *Staphylococcus aureus* and *Staphylococcus epidermidis* biofilms. *Antimicrob Agents Chemother*, 56: 5804-5810.
- Seguin, JC, Walker, RD, Caron, JP, Kloos, WE, George, CG, Hollis, RJ, Jones, RN & Pfaller, MA 1999, Methicillin-Resistant *Staphylococcus aureus* outbreak in a veterinary teaching hospital: Potential human-to-animal transmission. *Journal of Clinical Microbiology*, 37: 1459-1463.
- Tene, V, Malagón, O, Finzi, PV, Vidari, G, Armijos, C & Zaragoza, T 2007, An ethnobotanical survey of medicinal plants used in Loja and Zamora-Chinchi, Ecuador. *Journal of Ethnopharmacology*, 111: 63-81.
- Tsay, RW, Siu, LK, Fung, CP & Chang FY 2002, Characteristics of bacteremia between community-acquired and nosocomial *Klebsiella pneumoniae* infection: risk factor for mortality and the impact of capsular serotypes as a herald for community-acquired infection. *Archives of Internal Medicine*, 162: 1021-7.

Bibliographic information of this paper for citing:

Lattef Ismaeel, ZA 2024, Anatomical study with antibacterial and antibiofilm effects of *Erodium cicutarium* (L.) L'H phenolic roots extract. *Caspian Journal of Environmental Sciences*, 22: 137-146.
